

## Main sol. 119

KH <sub>2</sub> PO <sub>4</sub>	0.50	g
MgSO <sub>4</sub> x 7 H <sub>2</sub> O	0.40	g
NaCl	0.40	g
NH <sub>4</sub> Cl	0.40	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.05	g

## Trace element solution SL-10

Yeast extract (OXOID)	1.00	g
Na-acetate	1.00	g
Na-formate	2.00	g

## FeSO<sub>4</sub> x 7 H<sub>2</sub>O solution (0.1% w/v)

	2.00	ml
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## Sludge fluid

	50.00	ml
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## Fatty acid mixture

	20.00	ml
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Sodium resazurin (0.1% w/v)	0.50	ml
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NaHCO <sub>3</sub>	4.00	g
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L-Cysteine HCl x H <sub>2</sub> O	0.50	g
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Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.50	g
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Distilled water	930.00	ml
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1. Dissolve ingredients except bicarbonate, cysteine and sulfide. Sparge medium with 80% H<sub>2</sub> and 20% CO<sub>2</sub> gas mixture for 30 - 45 min to make it anoxic. Add and dissolve bicarbonate, adjust pH to 6.5 and dispense medium under 80% H<sub>2</sub> and 20% CO<sub>2</sub> gas atmosphere into anoxic Hungate-type tubes or serum vials to 30% of their volume and autoclave. Add cysteine and sulfide from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas. Prior to use check pH of complete medium and adjust to 6.8 - 7.0, if necessary.

2. Note: After growth has started and the culture is becoming turbid add sterile 80% H<sub>2</sub> and 20% CO<sub>2</sub> gas mixture to 0.5 - 1 bar overpressure.